## TWO NEW TAXA OF ORNITHOBOEA (GESNERIACEAE) FROM THAILAND

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The new species *Ornithoboea grandiflora* D.J. Middleton and new variety *Ornithoboea maxwellii* var. *minutiflora* D.J. Middleton are described.

Keywords. Limestone, Mae Hong Son, Tak.

#### INTRODUCTION

The genus *Ornithoboea* C.B. Clarke, in subfamily Didymocarpoideae, tribe Trichosporeae, subtribe Loxocarpinae, is one of the more instantly recognisable in Thai Gesneriaceae owing to its characteristic and complex corolla shape with the so-called circlet around the mouth of the corolla tube (see Fig. 1 in Scott & Middleton, 2014). It was only recently revised (Scott & Middleton, 2014) and 16 species were recognised, of which 11 occur in Thailand. On a collecting trip to north-western Thailand in October 2014, specimens of *Ornithoboea* were collected in Mae Hong Son and Tak provinces that appeared to belong to two taxa not included in Scott & Middleton (2014). This paper clarifies the status of this material, resulting in the description of one new species and one new variety, *Ornithoboea grandiflora* D.J.Middleton and *Ornithoboea maxwellii* var. *minutiflora* D.J. Middleton.

#### MATERIAL AND METHODS

The newly collected herbarium material was morphologically compared with material of existing species in BKF, E, K, and SING (Thiers, continuously updated), and to the species concepts in Scott & Middleton (2014). Fresh leaf fragments that had been dried and preserved in silica gel were also collected. The collection details and the herbaria where the vouchers are deposited are listed in Table 1.

The laboratory work was carried out at the Royal Botanic Garden Edinburgh and Singapore Botanic Gardens. Total genomic DNA extractions were made either following the cetyltrimethyl ammonium bromide method of Doyle & Doyle (1987) without further purification, or by using the innuPREP Plant DNA Kit (Analytik Jena,

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|  | Accessions |  |  |
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|   |  |   | GenBank accessions |           |
|---|--|---|--------------------|-----------|
| Taxon   | Collection data                              | Locality and date   | ITS                | trnL-trnF |
| Didissandra elongata (Jack)<br>C.B.Clarke subsp. minor<br>(Ridl.) A.Weber & B.L.Burtt | C. Puglisi et al. CP186 (BO, E)              | Indonesia, Sumatra, Bengkulu,<br>1 vii 2011                 | KP325420           | KP325427  |
| Didissandra sp.   | C. Puglisi et al. CP130 (BO, E)              | Indonesia, Sumatra, West<br>Sumatra, 24 vi 2011             | KP325422           | KP325429  |
| Kaisupeea herbacea<br>(C.B.Clarke) B.L.Burtt  | D.J. Middleton et al. 5625 (E)               | Thailand, Prachin Buri, Na Di,<br>20 viii 2012              | KU203832           | KU203927  |
| Kaisupeea herbacea<br>(C.B.Clarke) B.L.Burtt  | D.J. Middleton et al. 5282 (E)               | Thailand, Kanchanaburi, Sai<br>Yok, 7 viii 2012             | KU203831           | KU203926  |
| Kaisupeea herbacea<br>(C.B.Clarke) B.L.Burtt  | D.J. Middleton et al. 4518 (E)               | Thailand, Chiang Mai, Chom<br>Tong, 19 ix 2008              | KU203830           | KU203925  |
| Kaisupeea orthocarpa B.L.Burtt  | D.J. Middleton et al. 4356 (E)               | Thailand, Surat Thani,<br>Phanom, 7 ix 2008                 | KU203833           | KU203928  |
| Ornithoboea arachnoidea<br>(Diels) Craib  | <i>C. Puglisi</i> et al. LAOS114 (E, SING)   | Lao PDR, Luang Prabang, 1 x<br>2014                         | KY580817*          | KY580767* |
| Ornithoboea arachnoidea<br>(Diels) Craib  | <i>C. Puglisi</i> et al. LAOS152 (E, SING)   | Lao PDR, Luang Prabang, 2 x 2014                            | KY580818*          | KY580768* |
| Ornithoboea arachnoidea<br>(Diels) Craib  | D.J. Middleton et al. 5806<br>(BKF, E, SING) | Thailand, Mae Hong Son,<br>Muang Mae Hong Son, 21 x<br>2014 | KY580819*          | KY580769* |
| Ornithoboea arachnoidea<br>(Diels) Craib  | D.J. Middleton et al. 4538 (E)               | Thailand, Chiang Mai, Chiang<br>Dao, 20 ix 2008             | JN934709           | JN934751  |
| Ornithoboea arachnoidea<br>(Diels) Craib  | <i>D.J. Middleton</i> et al. 4523 (E)        | Thailand, Chiang Mai, Chiang<br>Dao, 20 ix 2008             | KY580820*          | KY580770* |

#### TABLE 1. (Continued)

#### GenBank accessions Collection data Locality and date ITS Taxon trnL-trnF KU203839 Ornithoboea barbanthera D.J. Middleton et al. 4257 Thailand, Prachuap Khiri KU203934 (BKF.E) Khan, Pran Buri, 4 ix 2008 **B.L.Burtt** Ornithoboea flexuosa (Ridl.) A.R. Rafidah FRI 64358 (KEP) Malaysia, Kedah, Gunung KU203836 KU203931 Keriang **B.L.Burtt** Ornithoboea grandiflora D.J. Middleton et al. 4975 Thailand, Mae Hong Son, KY580821\* KY580771\* D.J.Middleton (BKF, E) Pang Ma Pha, 23 xi 2009 Ornithoboea grandiflora D.J. Middleton et al. 5821 Thailand, Mae Hong Son, KY580823\* KY580773\* D.J.Middleton (BKF, E, SING) Pang Ma Pha, 22 x 2014 Ornithoboea grandiflora Thailand, Mae Hong Son, KY580772\* D.J. Middleton et al. 5826 KY580822\* (BKF, E, SING) Pang Ma Pha, 22 x 2014 D.J.Middleton Ornithoboea maxwellii D.J. Middleton et al. 5815 Thailand, Mae Hong Son, KY580825\* KY580775\* S.M.Scott var. maxwellii (BKF, E, SING) Muang Mae Hong Son, 21 x 2014 Thailand, Mae Hong Son, Mae Ornithoboea maxwellii D.J. Middleton et al. 5797 KY580824\* KY580774\* S.M.Scott var. maxwellii (BKF, E, SING) La Noi, 20 x 2014 Thailand, Tak, Mae Ramat, 19 Ornithoboea maxwellii D.J. Middleton et al. 5781 KY580826\* KY580776\* S.M.Scott var. minutiflora (BKF, E, SING) x 2014 **D**.LMiddleton Ornithoboea occulta B.L.Burtt D.J. Middleton et al. 5702 Thailand, Tak, Umphang, 15 x KY580827\* KY580777\* (BKF, E, SING) 2014 Thailand, Tak, Umphang, 18 x Ornithoboea occulta B.L.Burtt D.J. Middleton et al. 5770 KY580828\* KY580778\* (BKF, E, SING) 2014

TABLE 1. (Continued)

|   |   |   | GenBank accessions |           |
|---|---|---|--------------------|-----------|
| Taxon                                   | Collection data                                 | Locality and date   | ITS                | trnL-trnF |
| Ornithoboea pseudoflexuosa<br>B.L.Burtt | D.J. Middleton et al. 4426 (E,<br>KEP)          | Thailand, Trang, Huay Yot, 9<br>ix 2008                           | KY580829*          | KY580779* |
| Ornithoboea puglisiae<br>S.M.Scott      | D.J. Middleton et al. 5617<br>(BKF, E, P, SING) | Thailand, Nan, Muang Nan,<br>16 viii 2012                         | KU203840           | KU203935  |
| Ornithoboea puglisiae<br>S.M.Scott      | D.J. Middleton et al. 5814<br>(BKF)             | Thailand, Mae Hong Son,<br>Muang Mae Hong Son, 21 x<br>2014       | KY580830*          | KY580780* |
| Ornithoboea puglisiae<br>S.M.Scott      | D.J. Middleton et al. 5870<br>(BKF, SING)       | Thailand, Loei, Nong Hin, 2<br>viii 2015                          | KY582831*          | KY580781* |
| Ornithoboea puglisiae<br>S.M.Scott      | C. Puglisi et al. LAOS281 (E,<br>SING)          | Lao PDR, Luang Prabang,<br>Pha Tad Ke Botanic Garden,<br>9 x 2014 | KY580831*          | _         |
| Ornithoboea puglisiae<br>S.M.Scott      | P. Triboun 4627 (BK)                            | Thailand, Nan, Tham Pha<br>Tum, 7 ix 2011                         | KY580832*          | KY580782* |
| Ornithoboea wildeana Craib              | D.J. Middleton et al. 4531 (E)                  | Thailand, Chiang Mai, Chiang<br>Dao, 20 ix 2008                   | JN934752           | JN934710  |
| Ornithoboea wildeana Craib              | D.J. Middleton et al. 5000 (E)                  | Thailand, Chiang Mai, Chiang<br>Dao, 27 ix 2009                   | KY580833*          | KY580783* |
| <i>Rhabdothamnopsis sinensis</i> Hemsl. | <i>M. Möller &amp; P. Zhou</i> MMO 09–1613 (E)  | China, Sichuan, Luding, 17 viii<br>2009                           | KU203828           | KU203923  |

ITS, internal transcribed spacer.

\*Sequences newly published in this study. The other data were generated by the authors for earlier studies and previously uploaded on GenBank.

Jena, Germany) according to the manufacturer's instructions. The markers used in our analyses were the nuclear internal transcribed spacer (ITS), sequenced from the primer pairs 5P–8P (both Möller & Cronk, 1997) or ITS1–ITS4 (both White *et al.*, 1990), and the plastid *trnL-trn*F (i.e. *trnL* intron and *trnL-trn*F spacer), from the primer pair C–F (both Taberlet *et al.*, 1991). These are the regions that have been sequenced in other recent studies of Asian Gesneriaceae (Middleton *et al.*, 2015; Möller *et al.*, 2016; Puglisi *et al.*, 2016).

The PCRs followed two main 25- $\mu$ L recipes: (1) 2.5  $\mu$ L of 2 mM dNTPs, 2.5  $\mu$ L of  $10 \times \text{NH}_4$  buffer, 1.25  $\mu$ L of 25 mM MgCl<sub>2</sub>, 0.75  $\mu$ L each of 10  $\mu$ M forward and reverse primer, 0.8  $\mu$ L of 0.4% bovine serum albumin, 0.2  $\mu$ L of Biotaq polymerase (Bioline, London, UK), 1  $\mu$ L of DNA template and 15.25  $\mu$ L of dH<sub>2</sub>O; and (2) 12.5  $\mu$ L of GoTaq Green Master Mix (Promega, Madison, Wisconsin, USA), adding 1 µL each of 10  $\mu$ M forward and reverse primer, 1  $\mu$ L of DNA and 9.5  $\mu$ L of dH<sub>2</sub>O. The PCR thermocycles used were: (1, for some ITS) 94°C for 3',  $30 \times [94°C \text{ for } 1', 55°C \text{ for } 1']$ 72°C for 1.5'], 72°C for 5', 10°C forever; (2, for some *trn*L-*trn*F) 94°C for 4', 35 × [94°C for 45", 55°C for 45", 72°C for 3'], 72°C for 10', 10°C forever; and (3, for some ITS and *trnL-trnF*) 95°C for 3', 35 × [95°C for 1', 50°C for 1', 72°C for 1'], 72°C for 5', 4°C forever. The PCR products were purified with either ExoSAP-IT (Affymetrix, Santa Clara, California, USA), or QIAquick PCR Purification Kit (Qiagen, Redwood City, California, USA), from gel or directly from PCR product. All sequencing reactions used BigDye Terminator (Applied Biosystems, Thermo Fisher Scientific, Waltham, Massachusetts, USA) technology, and were sequenced by GenePool (University of Edinburgh, UK) or by AITbiotech (Singapore) on an ABI3730 DNA Analyser (Applied Biosystems). The use of different protocols and thermocycles was a practical choice based on the ready availability of chemicals in different laboratories, and not a form of troubleshooting. The sequences were edited in Sequencher version 4.7 (Gene Codes Corporation, Ann Arbor, Michigan, USA), manually aligned in Mesquite version 2.75 (Maddison & Maddison, 2011), and analysed in MrBayes version 3.2.2 (Ronquist & Huelsenbeck, 2003; Ronquist et al., 2011). The consensus trees were edited in FigTree version 1.3.1 (Rambaut & Drummond, 2009).

The outgroup was chosen to include *Kaisupeea* B.L.Burtt and *Rhabdothamnopsis* Hemsl., from the sister clade to *Ornithoboea* (Puglisi *et al.*, 2016), and accessions of *Didissandra* C.B.Clarke, which is part of Didissandrinae, the sister subtribe to the Loxocarpinae (Weber *et al.*, 2013). The ingroup includes samples of 8 of the 16 species recognised by Scott & Middleton (2014), plus the two putative new taxa (Table 1). All but one accession have full sequences of both DNA regions included; an accession of *Ornithoboea puglisiae* is represented only by the nuclear ITS and is therefore absent from the *trnL-trnF* consensus tree presented, although it is included in the combined analyses of the two markers.

The Bayesian analyses were run over 1,000,000 generations or longer at increments of 250,000 generations, until the standard deviation of the split sequences reached values lower than 0.01. Thus, the ITS Bayesian inference took 1,250,000 generations to reach a standard deviation of 0.009559, the *trnL-trn*F took 1,500,000 generations to

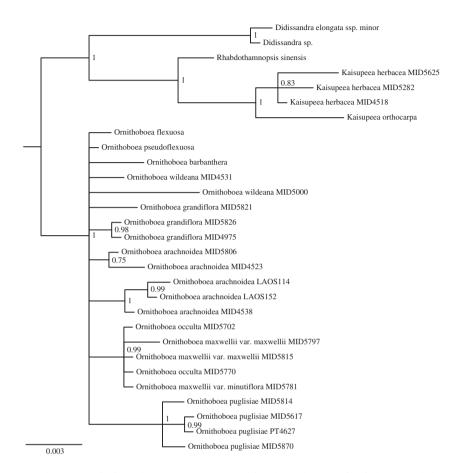


FIG. 1. The 50% majority rule consensus tree resulting from the Bayesian inference of the trnL-trnF data. Statistical support is expressed as posterior probabilities, which are shown by the nodes.

reach a standard deviation of 0.009080, and the combined dataset reached a standard deviation of 0.008579 after 1,000,000 generations. The other parameters used were constant for all the analyses. The heat was set at 0.05, because the default setting of 0.1 did not allow for satisfactory swaps between chains; the substitution model used was a GTR with gamma-distributed rate variation across sites, allowing for a proportion of invariable sites; and the burn-in was arbitrarily set to discard 25% of the trees. The output trees were summarised in 50% majority rule consensus trees.

#### Results

The phylogenetic trees (Figs 1–3) present a strongly supported (posterior probability, PP = 1) monophyletic *Ornithoboea*, with the ingroup accessions placed along a basal polytomy. As expected, given the low taxonomic level being investigated in this study,

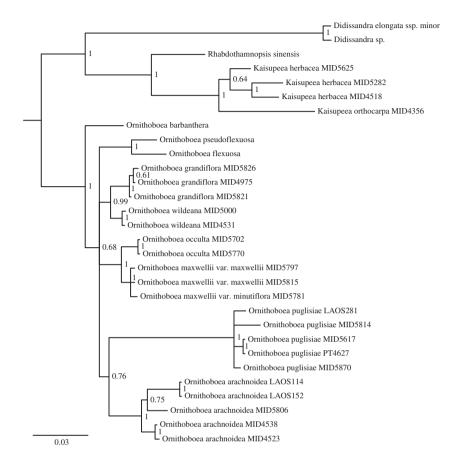


FIG. 2. The 50% majority rule consensus tree resulting from the Bayesian inference of the internal transcribed spacer data. Statistical support is expressed as posterior probabilities, which are shown by the nodes.

the *trnL-trn*F provided little insight into the relationships between species (Fig. 1). In the ITS tree, *Ornithoboea barbanthera* is sister to the rest of the genus (PP = 1), and the other accessions form four clades arranged in a polytomy. One of them sees *Ornithoboea pseudoflexuosa* as sister to *O. flexuosa*, with PP = 1. A second clade includes *Ornithoboea occulta*, *O. maxwellii* and the newly described *O. maxwellii* var. *minutiflora*. Support for the clade is high (PP = 1), as is the support for the monophyly of each species (PP = 1). A third clade includes *Ornithoboea wildeana* and the second putative new taxon, *O. grandiflora*; again, the clade and the monophyly of each of the two taxa receive very high support (PP = 0.99 and PP = 1, respectively). The last clade, supported by PP = 0.76, consists of the species *Ornithoboea puglisiae* and *O. arachnoidea*, both monophyletic with PP = 1 (Fig. 2).

Given the lack of topological conflict between the *trn*L-*trn*F and ITS tree, the datasets were also analysed together, under the same models but as distinct partitions,

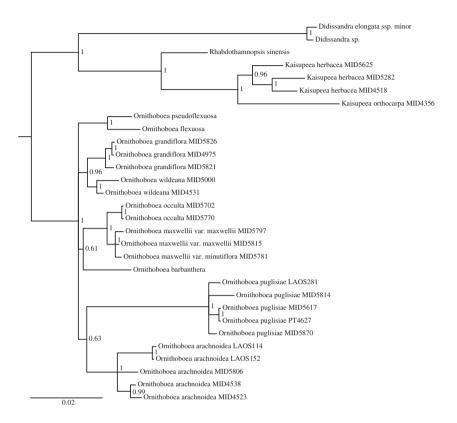


FIG. 3. The 50% majority rule consensus tree resulting from the Bayesian inference of a dataset combining internal transcribed spacer and *trnL-trnF* sequences. Statistical support is expressed as posterior probabilities, which are shown by the nodes.

in order to allow for overall different mutation rates across the regions. The resulting consensus tree (Fig. 3) presents one topological difference from the ITS tree, specifically in the position of *Ornithoboea barbanthera*, which is no longer sister to the whole genus, but rather is weakly associated with *O. occulta* and *O. maxwellii*, with PP = 0.61. The rest of the tree is otherwise topologically identical. The posterior probabilities of the nodes subtending *Ornithoboea wildeana* and *O. grandiflora* on one clade, and *O. puglisiae* and *O. arachnoidea* on another, are slightly lower (0.96 and 0.63, respectively), and some resolution is lost within *O. arachnoidea*.

#### DISCUSSION

The three accessions of the new species *Ornithoboea grandiflora*, identified on morphological grounds as distinct from *O. wildeana* and included in the molecular phylogenetic study, were found to form a monophyletic group sister to *O. wildeana*. *Ornithoboea maxwellii* var. *minutiflora*, identified on morphological grounds, was

initially thought distinct enough to be described as a new species, but on reflection and with the molecular phylogenetic study unable to distinguish it from *O. maxwellii* var. *maxwellii*, it was decided to describe it as a new variety. It should be noted, however, that the distinction of these new taxa was made primarily on morphological grounds, and that the molecular data do not refute these delimitations. Unfortunately, there was insufficient resolution to clarify the relationships between the species, which means it is not possible to test whether there are separate northern and southern clades, as found in *Paraboea* (Puglisi *et al.*, 2016).

### NEW TAXA

#### Ornithoboea grandiflora D.J.Middleton, sp. nov.

Most similar to *Ornithoboea wildeana* Craib and *Ornithoboea lacei* Craib in the large flowers, but differing from both in the geniculate filaments with no projection (large projection in *O. wildeana* and *O. lacei*); additionally differing from *O. wildeana* in the much denser pubescence in the palatal beard and circlet, and in the retuse corolla lobes; and differing from *O. lacei* in having smaller flowers. – Type: Thailand, Mae Hong Son, Pang Ma Pha, Tham Lod Cave area, 640 m, 22 x 2014, *D.J. Middleton, C. Hemrat, P. Karaket, C. Puglisi & S. Suddee* 5826 (holo BKF; iso E, QBG, SING [plus spirit]). Fig. 4.

Herb on limestone rocks; base of stem without persistent leaf bases; stems sparsely covered with long gland-tipped hairs, leaf internodes 4-6 cm. Leaves opposite; petioles densely short glandular pubescent and with scattered long gland-tipped hairs, (2.2–)5– 11.3 cm long; blade ovate,  $(3.5-)6.8-23 \times (1.6-)2.8-11.5$  cm, 1.4-2.6 times as long as wide, base cordate, apex acuminate, margin finely to coarsely crenate to dentate, 7-12 pairs of secondary veins, sparsely covered in a mixture of glandular hairs of various lengths above and beneath. Inflorescences cymose, axillary, 3.5–8 cm long, 4- to 14flowered, densely pubescent throughout with a mixture of shorter and longer glandular hairs; peduncle 1.2–2.8 cm long; bracts linear, 9–16 mm long; pedicels 8–19 mm long. Calyx of 5 lobes free to base, narrowly ovate, pinkish purple,  $9-12 \times 2.7-3.4$  mm, apex acuminate, sparsely to densely covered in short and long gland-tipped hairs. Corolla 16–19 mm long, tube, upper lip and tips of lobes on lower lip pale lilac to violet, rest of corolla cream-coloured, outside glabrous, inside densely pubescent on the circlet and at base of upper lip and lower lip (palatal beard), palatal beard pale yellow, hairs on circlet white, sessile glands dorsally in tube; tube 6.8–7.5 mm long; upper lip 2-lobed, 1-3 mm long, each lobe 3-4 mm wide; lower lip 3-lobed, not reflexed, 9.2-11.5 mm long, with distinct palatal beard inside at base, lateral lobes  $2.2-4.5 \times 3.7-4.6$  mm, mid lobe  $3.1-5.2 \times 3.7-4.5$  mm, all lobes oblong, apices retuse. *Stamens* inserted at c.3 mm from corolla base; filaments thin at base, thicker higher, thickened at bend but without large projection, strongly geniculate, densely pubescent at bend, c.5 mm long; anthers  $1.5-1.8 \times 2-2.6$  mm, densely pubescent at base, fused at tips; lateral staminodes 2.5–2.8 mm long, strongly curled downwards, medial staminode c.0.4 mm. Ovary 2.5– 3.2 mm long, densely covered in sessile to short-stalked glands; style 9-11 mm long,

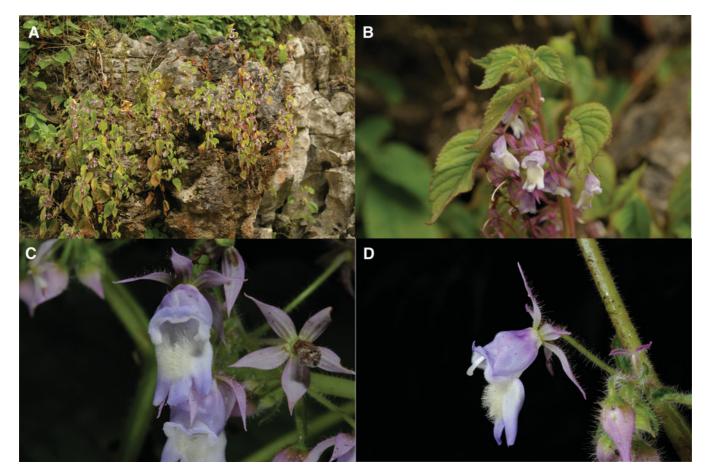


FIG. 4. Ornithoboea grandiflora D.J.Middleton. A, Habitat on limestone rocks; B, shoot and inflorescence; C, flower from the front; D, flower from the side. (A and B, photographs of *D.J. Middleton* et al. 5821 by Preecha Karaket; C and D, photographs of *D.J. Middleton* et al. 5826 by Preecha Karaket.)

sparsely to densely covered in subsessile to short-stalked glands, glabrous towards apex. *Stigma* truncate. *Fruit* unknown.

*Distribution*. Northern Thailand (known only from Mae Hong Son Province but close to the Myanmar border).

Habitat and ecology. In fissures in karst limestone rocks in full sun in dry evergreen forest.

*Etymology*. Named after the particularly large flowers. In the genus, only *Ornithoboea lacei* has larger flowers.

*Proposed IUCN conservation assessment.* Endangered (EN B1ab(ii,iii),B2ab(ii,iii)) under IUCN Standards and Petitions Subcommittee (2016) guidelines. The known extent of occurrence and area of occupancy, both less than 2 km<sup>2</sup>, place this in the Critically Endangered range owing to disturbance from human activity, especially where they occur along the road and as a result of construction of shrines. However, there is extensive limestone in the area, and the distribution could be greater than known. Also, at least two populations are known with no evidence of fragmentation, thereby qualifying the species as Endangered rather than Critically Endangered.

Additional specimens examined. THAILAND. Mae Hong Son: Pang Ma Pha, Road between Pai and Pang Ma Pha, 632 m alt., dry evergreen forest on limestone, 22 x 2014, *D.J. Middleton, C. Hemrat, P. Karaket, C. Puglisi & S. Suddee* 5821 (BKF, E, SING); Pang Ma Pha, Just outside entrance to Tham Lot cave area, 700 m alt., on large karst limestone boulder, 23 ix 2009, *D.J. Middleton, S. Lindsay & P. Suksathan* 4975 (BKF, E, QBG).

The large range in the lengths of the lobes in the lower lip of the corolla is a reflection of the degree of fusion of the central lobe to the lateral lobes, with shorter lobes corresponding to a longer fused part of the lower lip. It is highly variable even within a single plant.

Scott & Middleton (2014) included *Middleton* et al. 4975 in *Ornithoboea wildeana*, which is a widespread and variable species. The two species are superficially similar and can usually be distinguished by the slightly retuse corolla lobes in *Ornithoboea grandiflora*. *Ornithoboea grandiflora* has a denser pubescence in the palatal beard and circlet, but in the absence of plants of each species to compare directly, this is rather difficult to quantify for identification. The character that is most reliable to distinguish them is the lack of the filament projection in *Ornithoboea grandiflora* (a very pronounced filament projection in *O. wildeana*), but this requires dissection of a flower to see.

#### Ornithoboea maxwellii S.M.Scott var. minutiflora D.J.Middleton, var. nov.

Similar to *Ornithoboea maxwellii* S.M.Scott var. *maxwellii* but differing in the purple corolla lobes with purple hairs (white corolla lobes with white hairs or with very few purple hairs at base of lower lip in *O. maxwellii* var. *maxwellii*), and the uniformly minute hairs on the calyx (mixture of minute hairs and sparse long arachnoid hairs in *O. maxwellii* var. *maxwellii*). – Type: Thailand, Tak, Mae Ramat District, Road

# to Mae Sariang, 181 m alt., 19 x 2014, *D.J. Middleton, C. Hemrat, P. Karaket, C. Puglisi & S. Suddee* 5781 (holo BKF; iso E, QBG, SING [plus spirit]). Fig. 5.

Herb on limestone rocks; base of stems with persistent leaf bases; stems densely arachnoid pubescent (i.e. with long, twisting and matted hairs). Leaves opposite, crowded; petioles densely arachnoid pubescent, 2.7-11 cm long; blade ovate or orbicular,  $2.5-8.8 \times 2.2-7.8$  cm, 0.8-1.3 times as long as wide, base cordate, apex obtuse to rounded, margin coarsely crenate to dentate, 4-6 pairs of secondary veins, mixture of hairs of various lengths all over above, including sparse arachnoid hairs, similar beneath but more confined to venation so appearing sparser. Inflorescences cymose, 3- to 10-flowered, densely pubescent throughout with a mixture of shorter and longer glandular hairs and occasional much longer arachnoid hairs; peduncle 3.8–9 mm long; pedicels 5.5–10 mm long. Calyx of 5 lobes free to base, narrowly to broadly elliptic,  $3-4 \times 0.9-3.4$  mm, enlarging slightly and persistent in fruit, apex acute to acuminate, densely minutely puberulent. Corolla c.9.2 mm long, tube white, lobes purple, tube minutely and sparsely glandular puberulent, lobes with longer glandular hairs inside and outside, more densely so inside; tube c.3.8 mm long; upper lip 2lobed, each lobe  $1.8 \times 2.2$  mm, apices rounded, densely pubescent with purple hairs on upper lobes and in ring around throat; lower lip 3-lobed, not reflexed, c.5.4 mm long, lobes densely pubescent with purple hairs, without distinct palatal beard, lateral lobes c.2.8  $\times$  2.6 mm, mid lobe c.3.2  $\times$  2.5 mm, all lobes obovate, slightly overlapping, apices rounded. Stamens inserted at c.0.7 mm from corolla base; filaments thin at base, thickened higher, without pronounced sterile projection, geniculate, glabrous, c.2.2 mm long; anthers  $c.1 \times 1.2$  mm, glabrous, fused at the tips; staminodes 3, laterals c.1.5 mm long, medial c.0.1 mm long. Ovary c.1.5 mm long, densely glandular puberulent; style c.3 mm long, densely glandular puberulent. Stigma truncate. Fruit 7-8 mm long, 2-2.5 mm wide, densely long hairy, style persistent.

Distribution. Known only from the type locality.

*Habitat and ecology*. Limestone rocks and cliffs in mixed deciduous forest by road at 181 m altitude.

Etymology. Named after the small flowers, so far the smallest known in the genus.

*Proposed IUCN conservation assessment.* Data Deficient (DD) under IUCN Standards and Petitions Subcommittee (2016) guidelines. This species is currently known only from the type locality, which is not in a protected area. It is, however, in an area of extensive limestone that is not yet sufficiently explored and is also close to the border with Myanmar, which is even less well explored and where *Ornithoboea maxwellii* var. *minutiflora* could also occur.

This variety is similar to the recently described *Ornithoboea maxwellii* S.M.Scott var. *maxwellii* (Scott & Middleton, 2014), differing in the characters noted above and additionally, but rather more tentatively, in the corolla circlet being less hairy, the lower

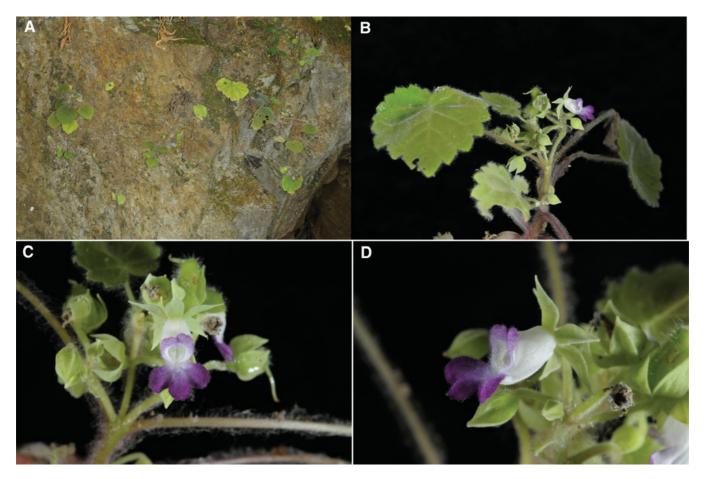


FIG. 5. Ornithoboea maxwellii var. minutiflora D.J.Middleton. A, Habitat on limestone rocks; B, whole plant; C, flower from the front; D, flower from the side. (Photographs of *D.J. Middleton* et al. 5781 by Preecha Karaket.)

lip rather less uniformly hairy, and the lower lobes shorter and wider. It should be noted for these latter characters, however, that *Ornithoboea maxwellii* var. *maxwellii* is known from only four collections and *O. maxwellii* var. *minutiflora* from only one. They are most easily distinguishable by the very obvious difference of *Ornithoboea maxwellii* var. *maxwellii* having white flowers, with an occasional purple hair at the base of the lower lip, and *O. maxwellii* var. *minutiflora* having the corolla limb purple, both in corolla tissue and hairs. In fruit, they can be distinguished by the indumentum on the persistent calyx, as noted above.

DNA sequence data show the two varieties to be almost identical for the two genes sequenced. Recognising that they are very close, but rather distinct from each other in the absence of any known intermediate morphologies, we describe this plant as a variety of *Ornithoboea maxwellii*.

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